

Research Article

Separate Effects of Ciprofloxacin, Metronidazole, or Minocycline on Microorganisms from Deep Caries

Divya Jha¹, Sagaw Prateepchinda², Helen H. Lu², and Gunnar Hasselgren^{1*}

¹Division of Endodontics, College of Dental Medicine, Columbia University, USA

²Department of Biomedical Engineering, Columbia University, USA

***Corresponding author**

Gunnar Hasselgren Division of Endodontics, College of Dental Medicine, Columbia University, 630 West 168 Street, New York NY 10032, USA; Tel: 1-212-305-0817; Fax: 1-212-305-8493; Email: bgh1@cumc.columbia.edu

Submitted: 14 June 2021

Accepted: 26 July 2021

Published: 28 July 2021

ISSN: 2333-7133

Copyright

© 2021 Jha D, et al.

OPEN ACCESS

Keywords

• Caries bacteria; Antibiotics; Ciprofloxacin; Metronidazole; Minocycline

Abstract

The study was carried out to evaluate antibacterial susceptibility of bacteria from deep carious lesions to 3 different antibiotics Ciprofloxacin, Metronidazole, or Minocycline that were tested at concentrations (0.5, 1, 2, 4, 8µg/ml) by the disk diffusion method. Carious, infected dentin close to the pulp was collected from teeth with deep cavities and cultured in thioglycolate and brain heart infusion broths for 48hrs anaerobically and aerobically.

The infected broths were tested for adequate growth and colony forming units against the McFarland Turbidity standard. It was then spread onto Brucella and Brain Heart Infusion Agar plates. Disks with the different concentrations were then placed on the agar surfaces. Each concentration was tested nine times. The control group consisted of agar plates infected with broth but without antibiotics. All agar plates were then placed in anaerobic or aerobic environments. The plates were checked for zones of inhibition after 48 hrs. All control groups had bacterial growth. A zone of inhibition was seen for all Ciprofloxacin concentrations tested under both aerobic and anaerobic conditions. Metronidazole and Minocycline did not show any zones of inhibition. Ciprofloxacin shows promise as an antibacterial agent against bacteria from deep caries.

INTRODUCTION

Pulp regeneration makes it possible to restore the vitality of an immature tooth and also to obtain root growth and increased dentin formation [1-5]. In order for this treatment to be successful freedom from infection is a prerequisite [6]. The most common source of pulp disease is caries and the flora from deep carious lesions is poly-microbial in nature and predominantly anaerobic [7-9]. When regenerative procedures are planned it is therefore necessary to use antibiotics with a wide spectrum to make sure that no microorganisms remain even after caries excavation. A combination of three antibiotics - Ciprofloxacin, Metronidazole, and Minocycline - has been introduced by Hoshino and collaborators [10] to be effective against bacteria stemming from caries. The three antibiotics mixture has been used for pulp regeneration [11-16] and also in pediatric patients against endodontic infections [17]. Only one of the three antibiotics, Metronidazole, has been tested separately for its effect on bacteria from caries [18,19]. Therefore, the aim of this study was to test the effects of all three antibiotics Ciprofloxacin, Metronidazole, and Minocycline separately on microorganisms from deep caries under anaerobic and aerobic conditions.

MATERIALS AND METHODS

Information about commercial products used in the study is summarized below. The study was approved by the Institutional Review Board of the Columbia University Irving Medical Center.

Cariou dentin was collected from nine teeth with deep carious lesions in seven patients (4 female, 3 male) ages 25 to 72. When the caries excavation came very close to the pulp the cavity, surrounding tooth structure, rubber dam and slow speed handpiece were wiped with small gauze soaked in 5% sodium hypochlorite solution and a new sterile round bur replaced the old one in the hand piece. Cariou dentin was collected from the flutes of the round bur after excavation of the deepest layer and transferred to Brain Heart Infusion and Thioglycolate broths. The mediums were incubated at 37° C in aerobic and anaerobic environments. After 48hrs the infected broths were used to infect fresh broths and also matched to the McFarland standard to approximate colony forming units. The infected broths were then plated on Brain Heart Infusion (BHI) or Brucella agar plates. Solutions of Ciprofloxacin, Metronidazole, or Minocycline were prepared at 0.5, 1, 2, 4, and 8µg/ml concentrations. The disk diffusion method according to the Clinical and Laboratory Standards Institute (Clinical and Laboratory Standards Institute, PO Box 633 Annapolis Junction, MD 20701 USA) was used to test the three antibiotics at the five different concentrations. Sterile disks were dipped into the different antibiotics concentrations and placed on the infected BHI or Brucella agar plates. Each concentration was tested nine times. All BHI plates were incubated aerobically at 37°C. Brucella plates were placed at 37°C in anaerobic bags with indicators to ensure an anaerobic environment was achieved. Eight plates, four BHI and four Brucella, were kept as controls. These were infected with the caries bacteria and no antibiotic

disks were placed on them. All plates were checked for a zone of inhibition after 48 hours. Statistical analysis: chi square test was used and the significance level was set at $P < 0.5$.

List of commercial products used in the study.

Product	Source
Brain Heart Infusion broth Systems, Morgan Hill, CA	AS872 Anaerobe
Thioglycolate broth Systems, Morgan Hill, CA	AS801 Anaerobe
Brain Heart Infusion agar plates Systems, Morgan Hill, CA	AS6424, Anaerobe
Brucella agar plates Systems, Morgan Hill, CA	AS111, Anaerobe
Sterile Disks Morgan Hill, CA	Anaerobe Systems,
McFarland standard Pleasant Prairie, WI	97298 MG Scientific,
Anaerobic bio-bags type A	BD Scientific, USA
Ciprofloxacin Switzerland	Fluka, Sigma-Aldrich,
Metronidazole Switzerland	Fluka, Sigma-Aldrich,
Minocycline	Sigma-Aldrich, USA

RESULTS

All control groups showed growth of bacteria. Ciprofloxacin showed zones of inhibition (Figure 1) at all concentrations in both aerobic and anaerobic environments. Metronidazole (Figure 2) and Minocycline did not show any zones of inhibition at any concentration in either aerobic or anaerobic environments. The differences in inhibition zones were found to be statistically significant $P = 0.007$.

DISCUSSION

The flora from deep carious lesions is poly-microbial and anaerobic bacteria dominate the flora [7-9]. Therefore it was

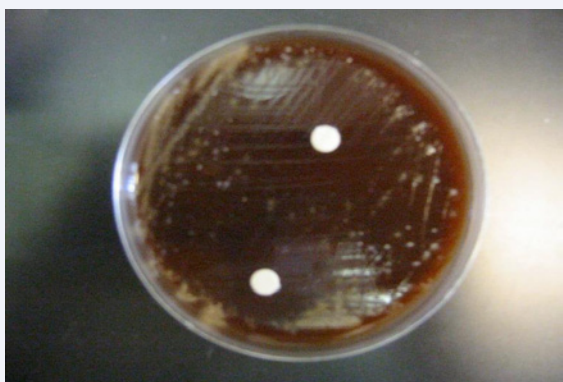


Figure 1 Ciprofloxacin 2µg/ml on brain heart infusion agar, clear zones of inhibition.



Figure 2 Metronidazole 8µg/ml on brucella agar, no zone of inhibition seen.

considered relevant to expose the three antibiotics to the whole caries flora both aerobically and anaerobically. The disk test of antibiotics sensitivity was used as it is a standard test for evaluating antibacterial activity [20]. The finding of no zones of inhibition around the test disks with Metronidazole or Minocycline does not mean that these antibiotics had no effect. A poly-microbial culture, contrary to a single species culture, is affected in multiple ways depending on the different species involved. Some of the microorganisms may have been killed or inhibited, whereas other species were unaffected and could therefore grow close to the test disk. Minocycline is a semi synthetic derivative of tetracycline having a broad spectrum of activity against gram positive and negative bacteria [21,22]. In addition to being an antibiotic Minocycline has other properties [23] including a wide use as an anti-inflammatory agent [24]. It has also been found that Minocycline inhibits angiogenesis [25] which would make it a less desirable drug when the goal is to achieve pulp regeneration. Metronidazole - 5-nitroimidazole - is an antibiotic that is active against most Gram-negative and Gram-positive anaerobic bacteria [26-28]. As deep caries contains mainly anaerobic microorganisms [9] Metronidazole would be expected to be effective against many caries bacteria. Earlier studies have found that metronidazole alone is efficient against caries bacteria both in vitro and in vivo [18,19]. The finding of this study that metronidazole did not eradicate all caries bacteria is likely to be caused by differences in testing techniques. Hoshino [19] enclosed Metronidazole in carious cavities which could add to the antibacterial effect of the antibiotic, whereas the present study used a disk diffusion method which does not have additional effects. Ciprofloxacin - fluoroquinolone - is a broad spectrum antibacterial agent. Most Gram-negative bacteria are highly susceptible to Ciprofloxacin, gram-positive bacteria are generally susceptible or moderately susceptible [29-31]. Ciprofloxacin showed clear zones of inhibition in both aerobic and anaerobic tests demonstrating that under the present conditions this antibiotic was capable of killing or suppressing the growth of all bacteria.

CONCLUSION

The results of the present study suggest that Ciprofloxacin alone can be the antibiotics of choice for eradication of bacteria from deep carious lesions.

REFERENCES

1. Nygaard-Ostby B, Hjortdal O. Tissue formation in the root canal following pulp removal. *Scand J Dent Res.* 1971; 79: 333-349.
2. Nevins AJ, Finkelstein F, Borden BG, Laporta R. Revitalization of pulpless open apex teeth in rhesus monkeys, using collagen-calcium phosphate gel. *J Endod.* 1976; 2: 159-165.
3. Nevins AJ, Cymerman JJ. Revitalization of Open Apex Teeth with Apical Periodontitis Using a Collagen-hydroxyapatite Scaffold. *J Endod.* 2015; 41: 966-973.
4. Bansal R, Jain A, Mittal S. Current overview on challenges in regenerative endodontics. *J Conserv Dent.* 2015; 18: 1-6.
5. Kim SG, Malek M, Sigurdsson A, Lin LM, Kahler B. Regenerative endodontics: a comprehensive review. *Int Endod J.* 2018; 51: 1367-1388.
6. Law AS. Considerations for regeneration procedures. *Pediatr Dent.* 2013; 35: 141-152.
7. Edwardsson S. Bacteriological studies on deep areas of carious dentine. *Odont Revy.* 25: 32. 1974.
8. Hoshino E. Predominant obligate anaerobes in human carious dentin. *J Dent Res.* 1985; 64: 1195-1198.
9. Siqueira JF. Pulpal infections, including caries. Chapter 10 in Seltzer and Bender's *Dental Pulp* 2nd ed. Eds Hargreaves KM, Goodis HE, Tay, FR. Quintessence 2012. 205-239.
10. Hoshino E, Kurihara-Ando N, Sato I, Uematsu H, Sato M, Kota K, et al. In-vitro antibacterial susceptibility of bacteria taken from infected root dentine to a mixture of ciprofloxacin, metronidazole and minocycline. *Int Endod J.* 1996; 29: 125-130.
11. Bose R, Nummikoski P, Hargreaves K. A retrospective evaluation of radiographic outcomes in immature teeth with necrotic root canal systems treated with regenerative endodontic procedures. *J Endod.* 2009; 35: 1343-1349.
12. Ding RY, Cheung GS, Chen J, Yin XZ, Wang QQ, Zhang CF. Pulp revascularization of immature teeth with apical periodontitis: a clinical study. *J Endod.* 2009; 35: 745-749.
13. Jung IY, Lee SJ, Hargreaves KM. Biologically based treatment of immature permanent teeth with pulpal necrosis: a case series. *Tex Dent J.* 2012; 129: 601-616.
14. Gelman R, Park H. Pulp revascularization in an immature necrotic tooth: a case report. *Pediatr Dent.* 2012; 34: 496-499.
15. Sönmez IS, Akbay Oba A, Erkmen Almaz M. Revascularization/Regeneration performed in immature molars: case reports. *J Clin Pediatr Dent.* 2013; 37: 231-234.
16. Phumpatrakom P, Srisuwan T. Regenerative capacity of human dental pulp and apical papilla cells after treatment with a 3-antibiotic mixture. *J Endod.* 2014; 40: 399-405.
17. Takushige T, Cruz EV, Asgor Moral A, Hoshino E. Endodontic treatment of primary teeth using a combination of antibacterial drugs. *Int Endod J.* 2004; 37: 132-138.
18. Hoshino E, Kota K, Sato M, Iwaku M. Bactericidal efficacy of metronidazole against bacteria of human carious dentin in vitro. *Caries Res.* 1988; 22: 280-282.
19. Hoshino E, Iwaku M, Sato M, Ando N, Kota K. Bactericidal efficacy of metronidazole against bacteria of human carious dentin in vivo. *Caries Res.* 1989; 23: 78-80.
20. Jorgensen JH, Ferraro MJ. Antimicrobial Susceptibility Testing: A Review of General Principles and Contemporary Practices. *Clin Infect Dis.* 2009; 49: 1749-1755.
21. Brogden RN, Speight TM, Avery GS. Minocycline: A Review of its Antibacterial and Pharmacokinetic Properties and Therapeutic Use. *Drugs.* 1975; 9: 251-291.
22. Bishburg E, Bishburg K. Minocycline--an old drug for a new century: emphasis on methicillin-resistant *Staphylococcus aureus* (MRSA) and *Acinetobacter baumannii*. *Int J Antimicrob Agents.* 2009; 34: 395-401.
23. Garrido-Mesa N, Zarzuelo A, Gálvez J. Minocycline: far beyond an antibiotic. *Br J Pharmacol.* 2013; 169: 337-352.
24. Clemens V, Regen F, Le Bret N, Heuser I, Hellmann-Regen J. Anti-inflammatory effects of minocycline are mediated by retinoid signaling. *BMC Neurosci.* 2018; 19: 58.
25. Yao JS, Shen F, Young WF, Yang GY. Comparison of doxycycline and minocycline in the inhibition of VEGF-induced smooth muscle cell migration. *Neurochem Int.* 2007; 50: 525-530.
26. Templeton R. Metabolism and pharmacokinetics of metronidazole. A review. In: *Metronidazole*. ed 2. Proceedings of the International Metronidazole Conference, Montreal, Quebec, Canada. Excerpta Medica, Amsterdam. 1977: 28-49.
27. Müller M. Mode of action metronidazole on anaerobic microorganisms. In: *Metronidazole*. ed 2. Proceedings of the Second International Symposium on Anaerobic Infections. 1979, Royal Society of Medicine. Grune & Stratton, Inc, New York. 1979: 223-228.
28. Goldman P. Metronidazole. *N Engl J Med.* 1980; 303: 1212-1218.
29. Gould IM, Milne K, Jason C. Concentration-dependent bacterial killing, adaptive resistance and post-antibiotic effect of ciprofloxacin alone and in combination with gentamicin. *Drugs Exper Clin Res.* 1990; 16: 621-728.
30. Davis R, Markham A, Balfour JA. Ciprofloxacin. An Updated Review of its Pharmacology, Therapeutic Efficacy and Tolerability. *Drugs.* 1996; 51: 1019-1074.
31. Campoli-Richards DM, Monk JP, Price A, Benfield P, Todd PA, Ward A. Ciprofloxacin. A review of its Antibacterial Activity, Pharmacokinetic Properties and Therapeutic Use. *Drugs.* 2012; 35: 373-447.

Cite this article

Jha D, Prateepchinda S, Lu HH, Hasselgren G (2021) Separate Effects of Ciprofloxacin, Metronidazole, or Minocycline on Microorganisms from Deep Caries. *JSM Dent* 9(1): 1136.